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In silico investigation of the anti-diabetic potential of garden egg compounds on alpha-glucosidase

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Abstract:

Diabetes mellitus is a chronic metabolic disorder affecting millions of people worldwide. In 2019, diabetes was the direct cause of 1.5 million deaths according to WHO. Alpha glucosidase is an intestinal enzyme that aids digestion of dietary carbohydrates and starches to produce glucose for intestinal absorption, which in turn leads to the increase in blood glucose levels. Hence, this study was aimed at investigating, through in silico approach, the possible inhibitory potential of Solanum aethiopicum natural product, solasodine, on alpha glucosidase through binding in the enzyme's active site. Thus, blocking its hydrolytic potential of cleaving oligosaccharides to absorbable monosaccharides. Following a preliminary screening of 149 ligands from PubChem database, seven (7) ligands were selected and their physiochemical and absorption, distribution, metabolism, elimination and toxicity (ADMET) properties were checked using Swiss institute bioinformatics webserver. Further molecular docking studies carried out revealed that Solasodine and Notoryne were the most potent inhibitors of alpha glucosidase with binding affinities of -10.05 kcal/mol and -7.50 kcal/mol, respectively. The findings of this study are believed to serve as a baseline information for further wet lab investigations by pharmaceutical organizations toward the development of new drugs for combating the endemicity of diabetes in this region of the globe.

Keywords: Garden egg, Solasodine, In silico, Diabetes, Solanum

1. Introduction

Diabetes Mellitus (DM) is a metabolic disorder characterized by elevated levels of blood glucose (hyperglycemia) resulting from either inadequate insulin production by the pancreas, insufficient cellular response to insulin or a combination of both factors [1]. It is a worldwide pandemic, ranking as the world's fifth leading cause of death. The International Diabetes Mellitus Federation predicts that 592 million people worldwide will have DM by 2035 [2]. As it stands, of the 537 million cases recorded globally in 2021, 24 million cases are from Africa; and Nigeria accounts for over 3 million cases [3]. Beyond its economic implications, DM adversely affects individuals' quality of life contributing to complications such as cardiovascular diseases, kidney failure and neuropathy [1]. In addition, DM exerts significant social and public health impact such as influencing employment opportunities, social relationships and mental health [3].

Alpha glucosidase is a membrane-bound enzyme situated in the epithelium of small intestine, playing a crucial role in enhancing glucose absorption, which is achieved by catalysing the hydrolytic cleavage of oligosaccharides into absorbable monosaccharides [4]. Consequently, the inhibition of this enzyme results in a deceleration of postprandial blood sugar elevation post-meal [5-8], and the

rate of carbohydrate digestion is reduced, thereby decreasing glucose absorption. Nonetheless, in diabetic patients, such therapies of enzyme inhibition have a short-term effect of lowering current blood glucose levels, with a slight long-term reduction in haemoglobin levels [4].

Solanum aethiopicum, known as "Garden egg" or "Scarlet eggplant," is a consumable vegetable crop categorized within the Solanaceae family. The Solanaceae family is one of the most extensive and vital groups of vegetables, primarily originating in tropical regions. Solanum aethiopicum is commonly cultivated in Nigeria for its nutritional and medicinal properties [9]. Several Solanum species have been reported to have anti-diabetic properties, making them promising plants for developing efficacious and safer medicines for diabetes and its complications [10-15]. Specific researches conducted on the phytochemical compositions of Solanum species have shown their abilities to stimulate insulin secretions [16] and lower high sugar diets [17]. Other functions include possessing insulin-like activities through their hypoglycaemic capabilities [18] and suppressing sucrose transfer to the intestines from the stomach [19].

While most studies have delved into the phytochemicals of this species, the present study was designed to investigate, *in silico*, the possible inhibitory

potential of a *Solanum aethiopicum* natural product, solasodine, on alpha glucosidase through binding in the enzyme's active site. Thus, blocking its hydrolytic potential of cleaving oligosaccharides to absorbable monosaccharides.

2. Materials and Methods

2.1 Protein sequence retrieval

The target protein, alpha glucosidase (5kzw), was retrieved from RSCB PDB database (http://www.rcsb.org/) [20].

2.2 Target protein preparation and active site prediction

Target protein was processed for docking by adding polar hydrogens, removing hetero-atoms, and then converting the data to ".pdbqt" file format as described earlier [21]. Furthermore, the active site of the targeted protein was predicted using CASTp calculations (Computed Atlas of Surface Topography of proteins) by going to the CASTp server and uploading the target protein pdb ID, then clicking on submit (http://sts.bioe.uic.edu/) [22].

2.3 Preparation of ligands for docking

Following a preliminary screening of 149 ligands from PubChem database (https://pubchem.ncbi.nlm.nih.gov/) [23], seven ligands were selected and docked to identify the best ligand with the least binding energy. The 3D structure of the ligands were downloaded from PubChem database in ".sdf" format and converted to ".pdbqt" format.

2.4 ADMET Analysis

ADMET properties of the selected ligands were checked using Swiss institute of bioinformatics webserver (https://www.sib.swiss/) [24]. The ligands were screened for drug likeness by their adherence to Lipinski's "rule of five", and with no more than one (1) violation; and their molecular properties [25].

2.5 Molecular docking using AutoDock

Each ligand was docked with alpha glucosidase protein using AutoDock 4.2.6 software (version 1.5.7). Lamarckian genetic algorithm (LGA) was used as a default search function. The energy value of the docked molecule was noted and calculated in kcal/mol from the output files. Similarly, inhibition constant (Ki) was predicted from the docking process. The binding site of the ligand was selected based on the active site component and the substratebinding site of the alpha glucosidase protein. The grid box size was set as Size X = 72 Å, Size Y = 66 Å and Size Z = 72 Å. The grid box centre was set as Centre X = - 0.558 Å, Centre Y = -41.04 Å and Centre Z = -18.486 Å. AutoGrid program supplied with AutoDock was used to produce grid map. The spacing between grid points was 0.375 Å. Docked conformations were clustered using a tolerance of 2.0 Å RMSD. Discovery software visualizer (Accelrys, Inc, USA) was employed to visualize and analyse the receptor and ligand structures.

3. Results and Discussion

Solasodine shows promising binding interactions to the target enzyme suggesting potential antidiabetic property. To investigate the inhibitory potential of garden egg compounds on alpha glucosidase *in silico*, seven compounds were docked to the enzyme via Autodock to demonstrate their binding potentials as shown in Table 1.

The table shows their docking score, inhibitory constant (Ki) values, ADMET properties, and interacting residues between the compounds and alpha glucosidase enzyme. It has been established from [25] that a more negative binding affinity indicates stronger binding. Similarly, Figure 1 shows the 2-D putative binding mode of the compounds with alpha glucosidase enzyme. Solasodine displayed the least binding energy score (-10.05 Kcal/mol) against alpha-glucosidase with six different amino acid residues.

Table 1	l: Selec	ted gar	den egg	compound	ds from F	PubChem	databa	se used	for o	docking
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Ligand	Docking Score (Kcal/mol)	Inhibition Ki (uM)	Lipinski rule of five	Log P	GI absorption	Interacting residues	Molecular weight (g/mol)	Common name
442985	-10.05	43.30	1 violation	4.67	High	Asp356, SO41025, Leu868,Val718, His717, Tyr360	413.64	Solasodine
23427873	-7.50	3.19	Yes	3.76	High	Glu196, Leu195, Pro198, Tyr360, Arg608, Asp365, Thr197, Leu355, Gly359, Val357, Val358	347.68	Notoryne
5281417	-7.28	4.63	Yes	0.62	Low	SO41025, Glu866, Met366, Arg608	340.28	Esculin
76330546	-6.99	7.56	Yes	2.57	High	Glu866, SO41025, His584, Phe362, Pro595, Leu865, Arg594, Val867, Met363, His717, Leu868	224.34	15- Norlubiminol

Ligand	Docking Score (Kcal/mol)	Inhibition Ki (uM)	Lipinski rule of five	Log P	GI absorption	Interacting residues	Molecular weight (g/mol)	Common name
442399	-6.78	10.72	Yes	3.61	High	GLu866, Leu865, His717, His584, Arg608, Arg594, Phe362, Pro595	218.33	Solavetivone
10036509	-6.64	13.67	Yes	2.61	High	Pro198, Leu195, Tyr360	222.32	Aethione
6072	-5.88	48.92	1 violation	0.88	Low	Glu196, His717, Tyr360, SO41025, Phe362, Arg608	436.41	Phlorizin

In this study, molecular docking effectively identified Solasodine as a potential alternative anti-diabetic compound through significant binding interactions to the target enzyme.

Decreasing the absorption of dietary energy molecules can be achieved by inhibiting the enzymes that are responsible for metabolism of carbohydrates, which mainly include alpha amylase and alpha glucosidase [26]. Therefore, one of the targets for the treatment of diabetes is

the inhibition of these enzymes activities [27]. Glycosidic linkages of α -D-(1, 4) in carbohydrates are cleaved by alpha amylase to produce oligosaccharides, which are further cleaved to monosaccharide glucose by alpha glucosidase. Therefore, inhibitors of these enzymes can delay the increase in blood glucose level in people who consume carbohydrate-rich food, and keep the postprandial hyperglycemia under control [27].

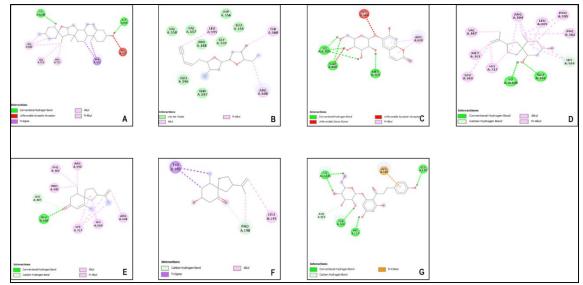


Figure 1: Putative binding mode of the seven garden egg compounds with alpha glucosidase. A: Solasodine; B: Notoryne; C: Esculine; D: 15-Norlubiminol; E: Solavetivone; F: Solavetivone; G: Phlorizin

These inhibitors are essentially other sugar molecules that compete with these enzymes. Alpha glucosidase inhibitors have been shown to effectively lower blood glucose levels by delaying the digestion and absorption of complex carbohydrates, improving blood sugar control in non-insulin-dependent diabetes mellitus patients [4]. Other alpha glucosidase inhibitors studied include commercial drugs such as acarbose, miglitol and voglibose [28]; and plant natural products such as Taxumariene F, Akebonoic acid, Morusin, Rhaponticin, Procyanidin A2, Alaternin, Mulberrofuran K and Psoralidin [5,29-30]. However, the use of these commercial drugs has imparted negative side effects such as abdominal distention and pain [31]. Notably, previous docking study with acarbose showed that it had a binding energy of -9.08Kcal/mol on lysozyme [32]. Therefore, the docking result obtained with Solasodine in this study with the biding energy of -10.05 kcal/mol suggests a possible better prospective anti diabetic drug on the alpha glucosidase enzyme. As determined through post docking

analysis, the molecular structure of *Solanum aethiopicum* natural compounds actively support hydrophobic interaction and form strong hydrogen bonds with adjacent amino acids. Consequently, this enhances the stability of the complex between the ligands and the enzyme, thereby increasing the effectiveness of enzyme inhibition.

Previous investigations have revealed the pharmacological activities associated with garden eggs, particularly their potential anti-ulcer [33], anti-oxidant [34], and anti-inflammatory properties [35-36]. Nonetheless, it is worth noting that this plant is often considered underutilized, possibly due to the limited knowledge and scientific information available about it [9]. Garden egg's composition is notable for its high crude fiber content, low fat content, and low dry matter, characteristics that may offer protection against various ailments such as constipation, colorectal carcinoma, diverticulitis, and atherosclerosis [37]. These attributes may also contribute to the observed weight reduction effects associated with the consumption of the

plant [37-38]. Additionally, garden egg boasts a noteworthy potassium content, an essential electrolyte that plays a vital role in maintaining cardiac function and regulating blood pressure. Others include iron, zinc, magnesium, sodium copper, manganese and calcium [39-40]. Furthermore, garden eggfruits contain polyphenols that inhibit key enzymes linked to type-2 diabetes and hypertension, potentially helping manage or prevent these degenerative conditions [39]. Other phytochemicals from *Solanum* spp studied to reduce postprandial glucose levels include phenols and polyphenolics [38-39], and sesquiterpenoids [41].

More so, general phytochemicals identified in *Solanum* spp include flavonoids, terpenoids, steroids, anthraquinones, tannins and phenols [40, 42]. Other phytochemicals with varying occurrence in *Solanum* spp include saponins and alkaloids [40].

4. Conclusion

In this study, the interaction between Solasodine and alpha glucosidase has offered valuable insight into molecular foundation that supports the established evidence regarding the antidiabetic attributes of the constituents of garden egg. Hence, the ability of this study to elucidate the relationship between binding affinities of the various biomolecules with respect to inhibition of alpha glucosidase could perhaps, serve as a baseline information in developing new antidiabetic drug.

References

- [1]. World Health Organization, (2021) Definition and diagnosis of diabetes mellitus and intermediate hyperglycemia: Report of a WHO/IDF consultation. World Health Organization.
- [2]. Guariguata, L., Whiting, D. R., Hambleton, I., Beagley, J., Linnenkamp, U. and Shaw, J. E., (2014). Global estimates of diabetes prevalence for 2013 and projections for 2035. *Diabetes Research* and Clinical Practice, 103(2), 137-149.
- [3]. International Diabetes Federation (2021). IDF Diabetes Atlas, 10th edn. Brussels, Belgium: (2021). Retrieved from https://idf.org/our-network/regions-and-members/africa/members/nigeria/
- [4]. Lebovitz, H., (1997). Alpha-Glucosidase inhibitors, Endocrinology and metabolism clinics of North America, 26(3), 539-551. https://doi.org/10.1016/S0889-8529(05)70266-8.
- [5]. Laar, F., Lucassen, P., Akkermans, R., Lisdonk, E., Rutten, G. and Weel, C., (2005). Alpha-glucosidase inhibitors for patients with type 2 diabetes: results from a Cochrane systematic review and metaanalysis. *Diabetes Care*, 28(1), 154-63. https://doi.org/10.2337/DIACARE.28.1.154.
- [6]. Yoshikawa, Y., Hirata, R., Yasui, H. and Sakurai, H., (2009). Alpha-glucosidase inhibitory effect of antidiabetic metal ions and their complexes. *Biochimie*, 91(10), 1339-1341. https://doi.org/10.1016/i.biochi.2009.06.005.

- [7]. Usman, B., Sharma, N., Satija, S., Mehta, M., Vyas, M., Khatik, G., Khurana, N., Hansbro, P., Williams, K. and Dua, K., (2019). Recent developments in alpha-glucosidase inhibitors for management of type-2 diabetes: an update. *Current Pharmaceutical Design*, 25(23), 2510-2525. https://doi.org/10.2174/1381612825666190717104 547.
- [8]. Zhang, X., Li, G., Wu, D., Yu, Y., Hu, N., Wang, H., Li, X. and Wu, Y., (2019). Emerging strategies for the activity assay and inhibitor screening of alphaglucosidase, Food and Function, 11(1), 66-82. https://doi.org/10.1039/c9fo01590f.
- [9]. Gruere, G., Giuliani, A. and Smale, M., (2006). Marketing underutilized plant species for benefit of the poor: a conceptual framework, *International Food Policy Research Institute*, *Washington, USA*.
- [10]. Gandhi, G., Ignacimuthu, S. and Paulraj, M., (2011). Solanum torvum Swartz. fruit containing phenolic compounds shows antidiabetic and antioxidant effects in streptozotocin induced diabetic rats, Food and Chemical Toxicology, An international journal published for the British Industrial Biological Research Association, 49(11), 2725-33. https://doi.org/10.1016/j.fct.2011.08.005.
- [11]. Sohrabipour, S., Kharazmi, F., Soltani, N. and Kamalinejad, M., (2013). Effect of the administration of *Solanum nigrum* fruit on blood glucose, lipid profiles, and sensitivity of the vascular mesenteric bed to phenylephrine in streptozotocin-induced diabetic rats *Medical Science Monitor Basic Research*, 19, 133 - 140. https://doi.org/10.12659/MSMBR.883892.
- [12]. Kandimalla, R., Kalita, S., Choudhury, B. and Kotoky, J., (2015). A review on anti-diabetic potential of genus Solanum (Solanaceae), Journal of Drug Delivery and Therapeutics, 5, 24-27. https://doi.org/10.22270/JDDT.V5I1.1026.
- [13]. Silva, E., Almeida-Lafetá, R., Borges, R. and Staerk, D., (2017). Dual high-resolution inhibition profiling and HPLC-HRMS-SPE-NMR analysis for identification of α-glucosidase and radical scavenging inhibitors in *Solanum americanum* Mill, *Fitoterapia*, 118, 42-48. https://doi.org/10.1016/j.fitote.2017.02.002.
- [14]. Umamageswari, M., Karthikeyan, T. and Maniyar, Y., (2017). Antidiabetic activity of aqueous extract of Solanum nigrum linn berries in alloxan induced diabetic wistar albino rats, Journal of Clinical and Diagnostic Research (JCDR), 11(7), FC16-FC19. https://doi.org/10.7860/JCDR/2017/26563.10312.
- [15]. Molehin, O., Elekofehinti, O. and Oyeyemi, A., (2020). Antihyperlipidemic, antiperoxidative and hypoglycemic effects of saponins from *Solanum* anguivi Lam: fruits in alloxan-induced diabetic rats. *The FASEB Journal*, 34(S1), 1-1. https://doi.org/10.1096/fasebj.2020.34.s1.00510.
- [16]. Pereira, A., Angolini, C., Souza-Sporkens, J., Silva, T., Oliveira, H. and Pastore, G., (2021). Brazilian sunberry (Solanum oocarpum Sendtn): Alkaloid composition and improvement of mitochondrial

- functionality and insulin secretion of INS-1E cells, Food Research International, 148, 110589. https://doi.org/10.1016/j.foodres.2021.110589.
- [17]. Nakitto, A., Rudloff, S., Borsch, C. and Wagner, A., (2021). Solanum anguivi Lam. fruit preparations counteract the negative effects of a high-sugar diet on the glucose metabolism in *Drosophila* melanogaster, Food and function, 12(19), 9238-9247.

https://doi.org/10.1039/d1fo01363g.

- [18]. Kar, D. M., Maharana, L., Pattnaik, S. and Dash, G. K., (2006). Studies on hypoglycaemic activity of Solanum xanthocarpum Schrad and Wendl. fruit extract in rats, Journal of Ethnopharmacology, 108(2), 251-256.
 - https://doi.org/10.1016/j.jep.2006.05.016
- [19]. Yoshikawa, M., Nakamura, S., Ozaki, K., Kumahara, A., Morikawa, T. and Matsuda, H., (2007). Structures of steroidal alkaloid oligoglycosides, robeneosides A and B, and antidiabetogenic constituents from the Brazilian medicinal plant *Solanum lycocarpum*, *Journal of Natural Products*, 70(2), 210-214. https://doi.org/10.1021/NP0680580.
- [20]. Berman, H. M., Westbrook, J., Feng, Z., Gilliland, G., Bhat, T. N., Weissig, H., Shindyalov, I. N. and Bourne, P. E. (2000). The protein data bank. *Nucleic Acids Research*, 28(1), 235-242. https://doi.org/10.1093/nar/28.1.235 in database: http://www.rcsb.org/
- [21]. Ja'afar, J. N., Ibrahim, A. and Zailani, H. A., (2023). *In silico* antimalarial investigation of some bioactive molecules targeting *Plasmodium falciparum's* sortilin protein, *Savannah Journal of Science and Engineering Technology*, 1(3), 50-54.
- [22]. Tian, W., Chen, C., Lei, X., Zhao, J. and Liang, J., (2018). CASTp 3.0: Computed atlas of surface topography of proteins. *Nucleic Acids Research*, 46(W1), W363-W367. https://doi.org/10.1093/nar/gky473 in database: http://sts.bioe.uic.edu/
- [23]. Kim, S., Chen, J., Cheng, T., Gindulyte, A., He, J., He, S., Li, Q., Shoemaker, B. A., Thiessen, P. A., Yu, B., Zaslavsky, L., Zhang, J. and Bolton, E. E. (2023). PubChem 2023 update. *Nucleic Acids Research*, 51(D1), D1373–D1380. https://doi.org/10.1093/nar/gkac956. In database https://pubchem.ncbi.nlm.nih.gov/
- [24]. Zoete, V., Daina, A., Bovigny, C. and Michielin, O., (2016). Swiss Similarity: A web tool for low to ultrahigh throughput ligand-based virtual screening. *Journal of Chemical Information and Modeling*, 56(8), 1399-1404. https://doi.org/10.1021/acs.jcim.6b00174 in database: https://www.sib.swiss/
- [25]. Lipinski, C. A., Lombardo, F., Dominy, B. W. and Feeney, P. J., (2012). Experimental and computational approaches to estimate solubility and permeability in drug discovery and development

- settings, Advanced Drug Delivery Reviews, 64, 4-17.
- [26]. Chia, C. W., Egan, J. M., and Ferrucci, L., (2018). Age-related changes in glucose metabolism, hyperglycemia, and cardiovascular risk, *Circulation Research*, 123(7), 886-904.
- [27]. Lordan, S., Smyth, T. J., Soler-Vila, A., Stanton, C. and Ross, R. P., (2013). The α-amylase and α-glucosidase inhibitory effects of Irish seaweed extracts, Food Chemistry, 141(3), 2170-2176.
- [28]. Dileep, K. V., Nithiyanandan, K. and Remya, C., (2018). Binding of acarbose, an anti-diabetic drug to lysozyme: a combined structural and thermodynamic study, *Journal of Biomolecular* Structure and Dynamics, 36(13), 3354-3361.
- [29]. Yamamoto, M. and Otsuki, M., (2006). Effect of inhibition of alpha-glucosidase on age-related glucose intolerance and pancreatic atrophy in rats, *Metabolism: Clinical and Experimental*, 55(4), 533-540.

https://doi.org/10.1016/J.METABOL.2005.11.007

[30]. Dirir, A., Daou, M., Yousef, A. and Yousef, L., (2021). A review of alpha-glucosidase inhibitors from plants as potential candidates for the treatment of type-2 diabetes, *Phytochemistry Reviews*, 21, 1049 - 1079.

https://doi.org/10.1007/s11101-021-09773-1.

- [31]. Cho, N. H., Shaw, J. E., Karuranga, S., Huang, Y., da Rocha Fernandes, J. D., Ohlrogge, A. W. and Malanda, B. I. D. F., (2018). IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and projections for 2045, *Diabetes Research and Clinical Practice*, 138, 271-281.
- [32]. Rasheed, L., Rehman, W., Rahim, F., Ali, Z., Alanazi, A. S., Hussain, R. and Ali Shah, S. A., (2023). Molecular modeling and synthesis of indoline-2, 3-dione-based benzene sulfonamide derivatives and their inhibitory activity against αglucosidase and α-amylase enzymes, ACS Omega, 8(17), 15660-15672.
- [33]. Chioma, A., Obiora, A. and Chukwuemeka, U., (2011). Does the African garden egg offer protection against experimentally induced ulcers. Asian Pacific Journal of Tropical Medicine, 4(2), 163-166.
- [34]. Umar, U. A., Hassan, L. G. and Maradun, K. L., (2019). TLC analysis and antioxidant activity of garden egg leaves, *Bayero Journal of Pure and Applied Sciences*, 12(1), 443-448.
- [35]. Anosike, C. A., Obidoa, O. and Ezeanyika, L. U., (2012). The anti—inflammatory activity of garden egg (*Solanum aethiopicum*) on egg albumin-induced oedema and granuloma tissue formation in rats, *Asian Pacific Journal of Tropical Medicine*, 5(1), 62-66.
- [36]. Olasunkanmi, A. A. and Afuye, O., (2021). Evaluation of Invitro Anti-Inflammatory Potential of Aqueous Solanum aethiopicum (Garden Egg) Leaf Extract, Further Biomedicines, 1(1), 1-14.
- [37]. Chinedu, S. N., Olasumbo, A. C., Eboji, O. K., Emiloju, O. C., Arinola, O. K. and Dania, D. I., (2011). Proximate and phytochemical analyses of

- Solanum aethiopicum L. and Solanum macrocarpon L. fruits, Research Journal of Chemical Sciences, 1(3), 63-71.
- [38]. Kwon, Y. I., Apostolidis, E. and Shetty, K. J. B. T., (2008). In vitro studies of eggplant (*Solanum melongena*) phenolics as inhibitors of key enzymes relevant for type 2 diabetes and hypertension, *Bioresource technology*, 99(8), 2981-2988.
- [39]. Nwanna, E., Ibukun, E., Oboh, G., Ademosun, A., Boligon, A. and Athayde, M., (2014). HPLC-DAD Analysis and In-Vitro Property of Polyphenols Extracts from (Solanum aethiopium) Fruits on α -Amylase, α -Glucosidase and Angiotensin - 1-Converting Enzyme Activities, International Journal of Biomedical Science (IJBS), 10, 272 - 281. https://doi.org/10.59566/ijbs.2014.10272
- [40]. Henry, M. U., Dogun, O., Ogenyi, R. A., Obidola, S. M. and Henry, U. I., (2022). Phytochemical, nutritional and trace element of some *Solanum* (Garden Egg), *Nigerian Journal of Biotechnology*, 39(2), 44-52.
- [41]. Nagaoka, T., Goto, K., Watanabe, A., Sakata, Y. and Yoshihara, T., (2001). Sesquiterpenoids in root exudates of *Solanum aethiopicum Zeitschrift für Naturforschung C*, 56(9-10), 707-713.
- [42]. Okafor, H. K., Odugbemi, A. I., Okezie, C. B. and Achebe, M. K., (2016). Antidiabetic and hypolipidaemic effects of garden egg (*Solanum aethiopicum*) leaf extract in beta-cells of streptozotocin induced diabetic male wistar rats, *Annual Research and Review in Biology*, 10(6), 1-